

PhosphoWorks™ Fluorimetric Pyrophosphate Assay Kit

Blue Fluorescence

Ordering Information

Product Number: 21611(2 plates)

Storage Conditions

Keep in freezer and protect from light

Instrument Platform

Fluorescence microplate readers

Introduction

Pyrophosphate (PPi) are produced by a number of biochemical reactions, such as ATP hydrolysis, DNA and RNA polymerizations, cyclic AMP formation by the enzyme adenylate cyclase and the enzymatic activation of fatty acids to form their coenzyme A esters.

Our PhosphoWorks™ Fluorimetric Pyrophosphate Assay Kit provides the most robust spectrophotometric method for the measurement of pyrophosphate. It uses our proprietary fluorogenic pyrophosphate sensor that has its fluorescence intensity proportionally dependent upon the concentration of pyrophosphate. Our assay is much easier and more robust than enzyme-coupling pyrophosphate methods, which require at least two enzymes for their pyrophosphate detections. Due to its direct measurement of pyrophosphate, this kit is ideal for screening inhibition or activities of enzymes that consume or generate pyrophosphate. The assay is an optimized mix-and-read assay and can be performed in a convenient 96-well or 384-well microtiter-plate format. The kit provides all the essential components for assaying pyrophosphate

Kit Key Features

Universal:	Can be used to monitor any biological processes that generate pyrophosphate.
Continuous:	Easily adapted to automation without mixing or separation.
Convenient:	Formulated to have minimal hands-on time.
Non-Radioactive:	No special requirements for waste treatment.

Kit Components

Components	Amount
Component A: Assay Buffer	1 bottle (25 mL)
Component B: PPi Sensor	1 vial (lyophilized powder)
Component C: Pyrophosphate Standard	1 mL (50 mM)
Component D: DMSO	1 vial (200 µL)

Assay Protocol (for one 96-well plate)

Brief Summary

Prepare pyrophosphate standards (50 µL) and/or test samples (50 µL) → Add Assay solution (50 µL) → Incubate at room temperature for 10 to 30 minutes
→ Monitor fluorescence intensity at Ex/Em = 316/456 nm

1. Prepare assay solution:

- 1.1 Thaw all the four components at room temperature before use.
- 1.2 Prepare 200X PPi Sensor Stock Solution: Add 50 µL of DMSO (Component D) into the vial of PPi Sensor (Component B) to make 200X PPi Sensor Stock Solution.

Note: 25 μ L of the PPi Sensor Stock Solution is enough for one 96-well plate. The unused PPi Sensor Stock Solution should be divided into single-use aliquots. Store at -20°C and protect from light.

- 1.3 **Prepare Assay Solution:** Add 25 μ L of 200X PPi Sensor Stock Solution (from Step 1.2) to 5 mL of Assay Buffer (Component A), and mix them well.

Note: Due to the high sensitivity of this assay to PPi, it is important to use PPi-free labware and reagents.

2. Prepare serially diluted pyrophosphate standards and test samples:

- 2.1 **Prepare 1 mM Pyrophosphate Standard Solution:** Add 10 μ L of 50 mM Pyrophosphate Standard (Component C) into 490 μ L of Assay Buffer (Component A), or buffer of your choice (preferably 50 mM Hepes buffer, pH 7) to make 1 mM pyrophosphate standard solution.

- 2.2 Add 50 μ L of 1 mM pyrophosphate standard solution (from Step 2.1) into 450 μ L of Assay Buffer (Component A) to get 100 μ M pyrophosphate standard solution, and then take 200 μ L of 100 μ M pyrophosphate standard solution to perform 1:3 serial dilutions to get 30, 10, 3, 1, 0.3, 0.1 and 0 μ M serially diluted pyrophosphate standards.

- 2.3 Add serially diluted pyrophosphate standards and/or pyrophosphate-containing test samples into a solid black 96-well microplate as described in Tables 1 and 2.

Table 1 Layout of pyrophosphate standards and test samples in a solid black 96-well microplate

BL	BL	TS	TS						
PS1	PS1						
PS2	PS2										
PS3	PS3										
PS4	PS4										
PS5	PS5										
PS6	PS6										
PS7	PS7										

Note: PS = Pyrophosphate Standard, BL = Blank Control, TS = Test Sample.

Table 2 Reagent composition for each well

Pyrophosphate Standards	Blank Control	Test Sample
Serial Dilutions*: 50 μ L	Assay Buffer: 50 μ L	50 μ L

*Note: *Add serially diluted pyrophosphate standards from 0.3 μ M to 100 μ M into wells from PS1 to PS7.*

3. Run pyrophosphate assay:

- 3.1 Add 50 μ L/well of Assay Solution (from Step 1.3) to the wells of pyrophosphate standards, blank control, and test samples. Mix the reagents thoroughly.

Note: For a 384-well plate, add 25 μ L of sample and 25 μ L of Assay Solution into each well.

- 3.2 Incubate at room temperature for 10 to 30 minutes.

Data Analysis

The fluorescence in blank wells (with the assay buffer only) is used as a control, and is subtracted from the values for those wells with the pyrophosphate reactions. A pyrophosphate standard curve is shown in Figure 1.

Note: The fluorescence background increases with time, thus it is important to subtract the fluorescence intensity value of the blank wells for each data point.

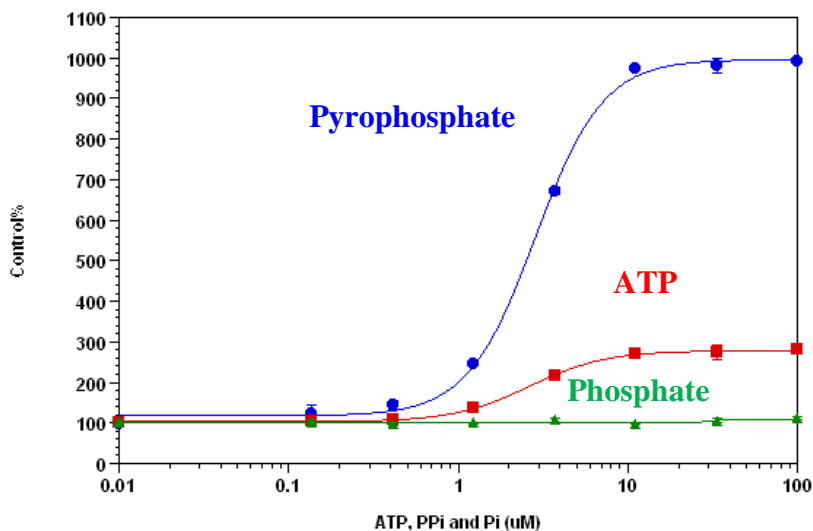


Figure1. Pyrophosphate, ATP and phosphate dose responses were measured with the PhosphoWorks™ Fluoremetric Pyrophosphate Assay Kit in a solid black 96-well plate using a fluorescence microplate reader. As low as 1 μ M (100 picomoles/well) pyrophosphate can be detected with 10 minutes incubation.

References

1. Zhou M, Diwu Z, Panchuk-Voloshina N and Haugland RP. (1997) A Stable Nonfluorescent Derivative of Resorufin for the Fluorometric Determination of Trace Hydrogen Peroxide: Applications in Detecting the Activity of Phagocyte NADPH Oxidase and Other Oxidases *Anal Biochem* 253, 162-168.
2. Mohanty, JG, Jaffe JS, Schulman E S and Raible DG. (1997) A highly sensitive fluorescent micro-assay of H_2O_2 release from activated human leukocytes using a dihydroxyphenoxazine derivative. *J. Immunol. Methods* 202: 133-141.

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